

Note

Implications of solvent selectivity triangles in assessing stationary phases for gas chromatography

T. J. BETTS

School of Pharmacy, Curtin University of Technology, GPO Box U1987, Perth, W.A. 6001 (Australia)
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Poole and Poole¹ have recently reviewed the characterization of solvent properties of gas chromatography (GC) liquid stationary phases. Referring to the use of Snyder's solvent selectivity triangle² which they note "has strong visual impact" they comment that this "original choice of (solvents for triangular) axes for proton-donor-acceptor and orientation interactions ... is quite reasonable". Snyder's probe solutes used for studies of GC stationary phases were ethanol, nitromethane (Y and U respectively as designated by Rohrschneider³) and dioxane (I' by McReynolds⁴) and Kersten and Poole⁵ in 1988 found that "the retention of ethanol and nitromethane on the non-polar GC phases is inadequate" even at their low temperature of 80.8°C. They also found that "dioxane is a rather insensitive probe for proton-donor interactions ... New probes should be selected". Poole thus ignores his own observations in the review¹ allowing him to reuse his previous triangular figures. The earlier paper⁵ comments that "*n*-butanol, nitropropane, 2-pentanone and pyridine (McReynolds' *y'*, *u'*, *z'* and *s'*, respectively) ... are better retained on most phases. These additional probes can be used to determine whether the choice of the test solute influences the position of a particular phase in the solvent selectivity triangle". As only three probes must be used to construct such a triangle, it seems self-evident that their choice must do so and I have examined this here. Kersten and Poole⁵ noted that (Snyder's) nitromethane does not behave characteristically of the other nitroalkanes on most selective (GC) phases. "The use of higher molecular weight nitroalkanes in place of nitromethane will significantly change the relative position of a phase in the selectivity triangle". They further comment "that both pyridine and 2-pentanone have reasonably large dipole moments compared to dioxane and consequently their retention will be more influenced by orientation interactions"⁵. It seems desirable to use these two solutes in solvent triangles.

Poole and Poole note my work⁶ using *n*-butanol, pyridine and 2-octyne (McReynolds' *k'*) for a selectivity triangle with the comment "it is not clear what advantage was gained by the change. 2-Octyne measures mainly dispersive interactions so that ... the triangle is changed"¹. It was chosen because, of the McReynolds solute probes, it gave the best spread of results in the triangle—surely a good reason? This allowed me to perceive three groups of stationary phases: (1) fully methyl polysiloxanes such as OV-1, SE-30, SP-2100; and phenyl-methyl (nominally 50:50)

polysiloxanes like OV-17 or SP-2250, with the sequence of probes being butanol-pyridine-octyne. (2) More polar polysiloxanes such as cyanopropyl-phenyl-methyl (25:25:50) like OV-225; and the trifluoropropyl-methyl (50:50) ones like OV-210, with the sequence of probes being butanol-octyne-pyridine. (3) Highly polar phases such as cyanopropyl-methyl (90:10) polysiloxanes like SP-2330; polyethylene glycols (PEG) (20M and 1500) and diethylene glycol succinate, with the sequence of probes being octyne-butanol-pyridine.

If such results can indicate that only three phases may be enough for most purposes—possibly three different polysiloxanes—surely this justifies the choice of solute probes?

Kersten and Poole⁵ detected but did not enclose four selectivity groups in their Snyder triangle (using the undesirable ethanol, nitromethane and dioxane), given below in my sequence and labelling: (A) phenyl-methyl (50:50) such as OV-17; and trifluoropropyl-methyl (50:50) polysiloxanes (QF-1). (B) Cyanopropyl-methyl (25:75?) polysiloxanes like the unusual OV-105. (C) Cyanopropyl-phenyl-methyl (25:25:50) polysiloxanes like OV-225; and polyethylene glycols (20 M). (D) Thiocyanates—synthesised by them; not widely used.

Apart from the thiocyanates, the other groups all show the same probe sequence of ethanol-nitromethane-dioxane, suggesting these solutes are a poor combination.

Although for some reason they did not study fully methyl polysiloxanes, nor Apiezon, nor diethylene glycol succinate they still failed to note what I previously pointed out⁶, that the most polar (thiocyanate) phases were nearest to the centre of the selectivity triangle, and the lower polarity groups A and B furthest from the centre. "A phase exhibiting minimum selectivity would be located at the centre of the triangle. The most selective phases are found towards the corners of the triangle"⁵. This can be achieved by applying bias to the triangle axes—Kersten and Poole use 0.2–0.7 for ethanol and nitromethane, but 0.1–0.6 for dioxane, these being not quite the same axis values as those of Snyder².

PROCEDURE

To check this possibility that strongly polar phases might be indicated triangularly to be less selective than those of low polarity, I plotted selectivity triangles using published McReynolds values⁴ for different combinations of three of his first five solutes. The combination of benzene-butanol-nitropropane gave poor discrimination as every GC phase yielded the probes in this sequence. Other combinations gave different degrees of discrimination and butanol-nitropropane-pyridine was selected. It is interesting that two of these solutes were probes I selected previously in 1986⁶.

Values for each of these probes were calculated by

$$x = (\overline{\text{McR}}_{\text{solute}}) / \Sigma(\overline{\text{McR}}_{\text{three solutes}})$$

where $\overline{\text{McR}}$ are the McReynolds values published⁴ and these are given in Table I and plotted in a biased triangle in Fig. 1.

DISCUSSION

It is clear in Fig. 1 that strongly polar phases are clustered together near to the centre of the selectivity triangle, which can be included with them in a circle of radius

TABLE I
 PROPORTIONS (x) OF THE SUM OF THREE RECORDED McREYNOLDS VALUES, OR FROM KERSTEN AND POOLE⁵

\overline{McR} = McReynolds value published⁴; PSX = polysiloxane; PEG = polyethyleneglycol. *x* Values in italics are in the range 0.327–0.335 (around the "mid" value of 0.333). $\overline{K/P}$ = Values calculated according to McReynolds procedure from data⁵; bu = butanol; di = dioxane; et = ethanol; nim = nitromethane; nip = nitropropane; py = pyridine.

No.	Stationary phase	<i>n</i> -Butanol		Nitropropane		Pyridine		$\overline{\Sigma McR}$		Probe solvent sequence
		\overline{McR}	<i>x</i>	\overline{McR}	<i>x</i>	\overline{McR}	<i>x</i>	$\overline{\Sigma McR}$	<i>x</i>	
1	Aptezon L grease	22	0.229	32	0.333	42	0.438	96		bu-nip-PY
2	PSX-fully methyl (SE-30, OV-1, SP-2100)	53	0.335	64	0.405	41	0.260	158		py-BU-nip
3	PSX-phenyl-methyl (50:50) (OV-17, SP-2250) ^a	158	0.262	243	0.403	202	0.335	603		bu-py-NIP
4	PSX-phenyl-methyl (75:25) (OV-25)	204	0.258	305	0.387	280	0.355	789		bu-py-NIP
5	PSX-trifluoropropyl-methyl (50:50) (OV-210, QF-1)	233	0.233	463	0.463	305	0.305	1001		bu-py-NIP
6	PSX-cyanopropyl-phenyl-methyl (2.5:2.5:50) (OV-225)	369	0.296	492	0.394	386	0.309	1247		bu-py-NIP
7	PEG 20M	536	0.337	572	0.353	510	0.315	1618		py-BU-nip
8	PSX-cyanopropyl-phenyl (50:50) (SP-2300)	495	0.299	637	0.383	531	0.319	1663		bu-py-NIP
9	Free fatty acid phase	580	0.321	602	0.333	627	0.347	1809		bu-nip-PY
10	PEG 1500	607	0.333	626	0.344	589	0.323	1822		py-BU-nip
11	Diethylene glycol succinate	751	0.306	840	0.343	860	0.351	2451		bu-nip-PY
12	Tris-(cyano-ethoxy)propane	857	0.306	1028	0.367	915	0.327	2800		bu-py-NIP
13	Bis-(cyano-ethyl)formamide	991	0.320	1110	0.358	1000	0.322	3101		bu-py-NIP
		<i>Ethanol</i>		Nitromethane		Dioxane		$\overline{\Sigma K/P}$		
		$\overline{K/P}$	<i>x</i>	$\overline{K/P}$	<i>x</i>	$\overline{K/P}$	<i>x</i>	$\overline{\Sigma K/P}$		
3'	PSX-phenyl-methyl (50:50)	139	0.238	263	0.450	182	0.312	584		et-di-NIM
5'	PSX-trifluoropropyl-methyl (50:50)	263	0.230	427	0.484	253	0.286	883		et-di-NIM
6'	PSX-cyanopropyl-phenyl-methyl (2.5:2.5:50)	383	0.311	517	0.419	333	0.270	1233		di-ET-nim
7'	PEG 20M	581	0.330	727	0.413	450	0.256	1758		di-ET-nim

^a Approximate side chain percentage is indicated.

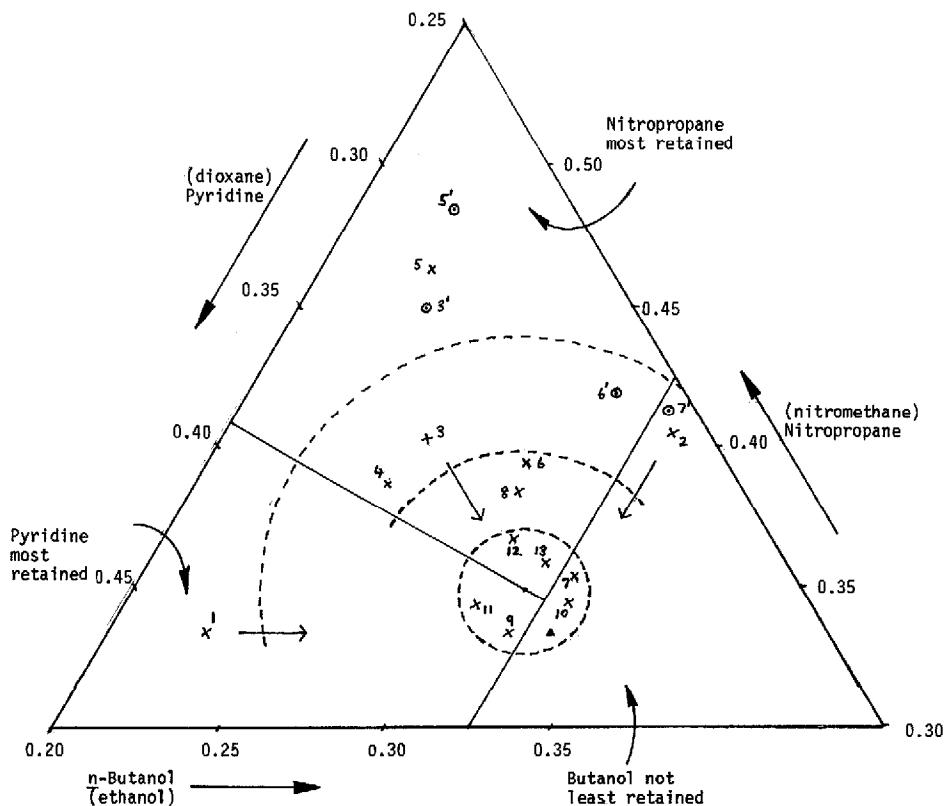


Fig. 1. Plot of McReynolds values on various gas chromatographic stationary phases (\times) for the three solute probes *n*-butanol (from left side to lower right apex), nitropropane (top apex) and pyridine (lower left apex). Also shown are values from Kersten and Poole⁵ for corrected I_R (retention index based on *n*-alkanes) on some of the phases (\circ) for the solutes ethanol (lower right apex), nitromethane (top apex) and dioxane (lower left apex). Numbers 1–13 refer to the phases listed in Table I. \blacktriangle is the centre of the full triangle plot (0.333 for all three solutes).

about 0.02 drawn at about 0.318 butanol–0.333 pyridine. Phases of fairly high polarity (SP-2300 and OV-225) are outside this circle whilst lower polarity SE-30, OV-17 and OV-25 are more remote. The most distant phases are Apiezon L and OV-210, and these are directed towards the pyridine-retaining and nitropropane-retaining corners of the triangle, respectively. It is possible to draw a line at butanol 0.325 to include the phases SE-30 and polyethylene glycols for which butanol is not the least retained (probe sequence pyridine–butanol–nitropropane). Another line separates from the remaining phases three which retain pyridine most strongly (diethylene glycol succinate, free fatty acid phase and Apiezon L) with probe sequence butanol–nitropropane–pyridine. This leaves a large group with strong affinity for nitropropane and least retention of butanol (probe sequence butanol–pyridine–nitropropane). These solute sequence-defined groups are very like my 1986 ones⁶, with nitropropane in place of octyne.

These divisions are quite odd, as each includes both low and high polarity phases. For example, fully methyl polysiloxane is grouped with the polyethylene

glycols, and Apiezon L is grouped with a polyester. What are the implications? It cannot be suggested sensibly that highly polar phases such as free fatty acid phase and PEG 1500 should be discarded as lacking solute discrimination because they plot near the 0.333 centre of the triangle. Nor is it reasonable to suggest that only phases near the apices of the triangle (Apiezon L and OV-210) should be used for their high discrimination. Perhaps phases should be selected for particular separations if they are virtually "neutral" to one of the solute probes (with a value close to 0.333) and hence preferentially retain one of the other two, so discriminating well, substances of this type retained? For example, Apiezon L (1 in Fig. 1) is neutral to nitroalkanes and selective for aromatics like pyridine, which it retains relatively strongly. It should not be chosen to resolve aliphatic alcohols. Free fatty acid phase (9) shows this to much lesser extent. Fully methyl polysiloxanes (2) are neutral to these alcohols, selective for nitroalkanes, and should not be chosen for aromatics; and this applies to PEG 1500 (10) to a lower degree. The commonly used phenyl-methyl (50:50) polysiloxanes (3) are surprisingly neutral to aromatics, selective for nitroalkanes, but not the phase of choice for aliphatic alcohols. Without exhibiting any fully "neutral" character tris-(cyano-ethoxy)propane (12) shows a reduced response of this type. Of course, this reduced discrimination response is due partly to the larger figures of McReynolds values given by the polar phases. Differences between the smaller values of the non-polar phases produce a relatively more severe displacement from the centre of the triangle.

Perhaps the best use for this plot is to indicate, for example, that Apiezon L is a low polarity alternative to diethylene glycol succinate. Similarly tris-(cyano-ethoxy)propane, OV-225, OV-17, OV-210 represent a decreasing polarity sequence of similar discriminatory potential. Corresponding results are obtained (with a somewhat different plot) using pentanone values with butanol and pyridine. Even using Kersten and Poole's results⁵ for ethanol-nitromethane-dioxane and plotting them on my butanol-nitropropane-pyridine triangle gives points for the phases which remain in the same groupings (Fig. 1).

The British Pharmacopoeia 1988 (Vol. I)⁷ includes over 60 examples of the use of GC on liquid phases and hence should provide a useful review of the selection of stationary phases for drug analysis. From my pyridine-retaining group, Apiezon L is only recommended twice, free fatty acid phase three times and polyester phases eight times. These phases thus form only about 22% of the GC uses. From my butanol-not-least-retained group, polyethylene glycols are recommended for twelve uses and fully methyl polysiloxanes for thirteen, giving 42% of the British Pharmacopoeia analyses and forming the main group. Most of the 36% remaining are from the butanol-pyridine-nitropropane group, although trifluoropropyl-methyl polysiloxanes such as OV-210 are not used at all. Perhaps these phases and Apiezon L, should be utilised?

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